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(54) Title: THERMOSTABLE DNA POLYMERASES		
<p>(57) Abstract</p> <p>An enzymatically active DNA polymerase having between 540 and 582 amino acids having a tyrosine at a position equivalent to position 667 of Taq DNA polymerase, wherein said polymerase lacks 5' to 3' exonuclease activity, and wherein said polymerase has at least 95 % homology in its amino acid sequence to the DNA polymerase of <i>S(i)(Thermus aquaticus)</i>, <i>S(i)(Thermus flavus)</i> or <i>S(i)(Thermus thermophilus)</i>, and wherein said polymerase forms a single polypeptide band on an SDS PAGE.</p>		

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DESCRIPTIONThermostable DNA polymerasesBackground of the Invention

The present invention relates to novel thermostable DNA polymerases, the genes and vectors encoding them and their use in DNA sequencing.

- 5 US Patents 4,889,818 and 5,079,352 describe the isolation and expression of a DNA polymerase known as Taq DNA Polymerase (hereinafter referred to as Taq). It is reported that amino-terminal deletions wherein approximately one-third of the coding sequence is absent  
10 have resulted in producing a gene product that is quite active in polymerase assays. Taq is described as being of use in PCR (polymerase chain reaction).

US Patent No. 5,075,216 describes the use of Taq in DNA sequencing.

- 15 International patent application WO 92/06/06188 describes a DNA polymerase having an identical amino acid sequence to Taq except that it lacks the N-terminal 235 amino acids of Taq and its use in sequencing. This DNA polymerase is known as  $\Delta$  Taq.

- 20 US Patent 4,795,699 describes the use of T7 type DNA polymerases (T7) in DNA sequencing. These are of great use in DNA sequencing in that they incorporate dideoxy nucleoside triphosphates (NTPs) with an efficiency comparable to the incorporation of deoxy  
25 NTPs; other polymerases incorporate dideoxy NTPs far less efficiently which requires comparatively large

quantities of these to be present in sequencing reactions.

At the DOE Contractor-Grantee Workshop (Nov. 13-17, 1994, Santa Fe) and the I. Robert Lehman Symposium (Nov 11-14, 1994, Sonoma), Prof. S. Tabor identified a site in DNA polymerases that can be modified to incorporate dideoxy NTPs more efficiently. He reported that the presence or absence of a single hydroxy group (tyrosine vs. phenylalanine) at a highly conserved position on *E. coli*, DNA Polymerase I, T7, and Taq makes more than a 1000-fold difference in their ability to discriminate against dideoxy NTPs. (See also European Patent Application 94203433.1 published May 31, 1995, Publication No. 0 655 506 A1 and hereby incorporated by reference herein.)

#### Summary of the Invention

The present invention provides a DNA polymerase having an amino acid sequence differentiated from Taq in that it lacks the N-terminal 272 amino acids and has the phenylalanine at position 667 (of native Taq) replaced by tyrosine. Preferably, the DNA polymerase has methionine at position 1 (equivalent to position 272 of Taq) (hereinafter referred to as FY2). The full DNA sequence is given as Fig 1 (SEQ. ID. NO. 1). Included within the scope of the present invention are DNA polymerases having substantially identical amino acid sequences to the above which retain thermostability and efficient incorporation of dideoxy NTPs.

By a substantially identical amino acid sequence is meant a sequence which contains 540 to 582 amino acids that may have conservative amino acid changes compared with Taq which do not significantly influence thermostability or nucleotide incorporation, i.e. other than the phenylalanine to tyrosine conversion. Such changes include substitution of like charged amino acids for one another, or amino acids with small side chains for other small side chains, e.g., ala for val. More drastic changes may be introduced at noncritical regions where little or no effect on polymerase activity is observed by such a change.

The invention also features DNA polymerases that lack between 251 and 293 (preferably 271 or 272) of the N-terminal amino acids of *Thermus flavus* (Tfl) and have the phenylalanine at position 666 (of native Tfl) replaced by tyrosine; and those that lack between 253 and 295 (preferably 274) of the N-terminal amino acids of *Thermus thermophilus* (Tth) and have the phenylalanine at position 669 (of native Tth) replaced by tyrosine.

By efficient incorporation of dideoxy NTPs is meant the ability of a polymerase to show little, if any, discrimination in the incorporation of ddNTPs when compared with dNTPs. Suitably efficient discrimination is less than 1:10 and preferably less than 1:5. Such discrimination can be measured by procedures known in the art.

One preferred substantially identical amino acid sequence to that given above is that which contains 562 amino acids having methionine at position 1 and alanine

at position 2 (corresponding to positions 271 and 272 of native Taq) (hereinafter referred to as FY3). A full DNA sequence is given as Fig. 2. This is a preferred DNA polymerase for expression by a gene of the present invention.

The purified DNA polymerases FY2 and FY3 both give a single polypeptide band on SDS polyacrylamide gels, unlike  $\Delta$  Taq, having either a phenylalanine or tyrosine at position 667 which forms several polypeptide bands of similar size on SDS polyacrylamide gels.

A second preferred substantially identical amino acid sequence is that which lacks 274 of the N-terminal amino acids of Thermus thermophilus having methionine at position 1, and the phenylalanine to tyrosine mutation at position 396 (corresponding to position 669 of native Tth) (hereinafter referred to as FY4). A full DNA sequence is given as Fig. 5 (SEQ. ID. NO. 14).

The present invention also provides a gene encoding a DNA polymerase of the present invention. In order to assist in the expression of the DNA polymerase activity, the modified gene preferably codes for a methionine residue at position 1 of the new DNA polymerase. In addition, in one preferred embodiment of the invention, the modified gene also codes for an alanine at position 2 (corresponding to position 272 of native Taq).

In a further aspect, the present invention provides a vector containing the gene encoding the DNA polymerase activity of the present invention, e.g., encoding an amino acid sequence differentiated from native Taq in that it lacks the N-terminal 272 amino acids and has

phenylalanine at position 396 (equivalent to position 667 of Taq) replaced by tyrosine or a substantially identical amino acid sequence thereto.

In a yet further aspect, the present invention provides a host cell comprising a vector containing the gene encoding the DNA polymerase activity of the present invention, e.g., encoding an amino acid sequence differentiated from native Taq in that it lacks the N-terminal 272 amino acids and has phenylalanine at position 396 (equivalent to position 667 of native Taq) replaced by tyrosine or a substantially identical amino acid sequence thereto.

The DNA polymerases of the present invention are preferably in a purified form. By purified form is meant that the DNA polymerase is isolated from a majority of host cell proteins normally associated with it; preferably the polymerase is at least 10% (w/w) of the protein of a preparation, even more preferably it is provided as a homogeneous preparation, e.g., a homogeneous solution. Preferably the DNA polymerase is a single polypeptide on an SDS polyacrylamide gel.

The DNA polymerases of the present invention are suitably used in sequencing, preferably in combination with a pyrophosphatase. Accordingly, the present invention provides a composition which comprises a DNA polymerase of the present invention in combination with a pyrophosphatase, preferably a thermostable pyrophosphatase such as *Thermoplasma acidophilum* pyrophosphatase. (Schafer, G. and Richter, O.H. (1992) Eur. J. Biochem. 209, 351-355).

The DNA polymerases of the present invention can be constructed using standard techniques. By way of example, mutagenic PCR primers can be designed to incorporate the desired Phe to Tyr amino acid change (FY mutation) in one primer. In our hands these primers also carried restriction sites that are found internally in the sequence of the Taq polymerase gene clone of Delta Taq, pWB253, which was used by us as template DNA. However, the same PCR product can be generated with this primer pair from any clone of Taq or with genomic DNA isolated directly from *Thermus aquaticus*. The PCR product encoding only part of the gene is then digested with the appropriate restriction enzymes and used as a replacement sequence for the clone of Delta Taq digested with the same restriction enzymes. In our hands the resulting plasmid was designated pWB253Y. The presence of the mutation can be verified by DNA sequencing of the amplified region of the gene.

Further primers can be prepared that encode for a methionine residue at the N-terminus that is not found at the corresponding position of Taq, the sequence continuing with amino acid residue 273. These primers can be used with a suitable plasmid, e.g., pWB253Y DNA, as a template for amplification and the amplified gene inserted into a vector, e.g., pRE2, to create a gene, e.g., pRE273Y, encoding the polymerase (FY2). The entire gene can be verified by DNA sequencing.

Improved expression of the DNA polymerases of the present invention in the pRE2 expression vector was obtained by creating further genes, pREFY2pref (encoding



a protein identical to FY2) and pREFY3 encoding FY3. A mutagenic PCR primer was used to introduce silent codon changes (i.e., the amino acid encoded is not changed) at the amino terminus of the protein which did not affect the sequence of the polypeptide. These changes led to increased production of FY2 polymerase. FY3 was designed to promote increased translation efficiency *in vivo*. In addition to the silent codon changes introduced in pREFY2pref, a GCT codon was added in the second position (SEQ. ID. NO. 2), as occurs frequently in strongly expressed genes in *E. coli*. This adds an amino acid to the sequence of FY2, and hence the protein was given its own designation FY3. Both constructs produce more enzyme than pRE273Y.

Silent codon changes such as the following increase protein production in *E. coli*:

- substitution of the codon GAG for GAA;
- substitution of the codon AGG, AGA, CGG or CGA for CGT or CGC;
- substitution of the codon CTT, CTC, CTA, TTG or TTA for CTG;
- substitution of the codon ATA for ATT or ATC;
- substitution of the codon GGG or GGA for GGT or GGC.

The present invention also provides a method for determining the nucleotide base sequence of a DNA molecule. The method includes providing a DNA molecule annealed with a primer molecule able to hybridize to the DNA molecule; and incubating the annealed molecules in a vessel containing at least one deoxynucleotide triphosphate, and a DNA polymerase of the present invention. Also provided is at least one DNA synthesis

terminating agent which terminates DNA synthesis at a specific nucleotide base. The method further includes separating the DNA products of the incubating reaction according to size, whereby at least a part of the  
5 nucleotide base sequence of the DNA molecule can be determined.

In preferred embodiments, the sequencing is performed at a temperature above 50°C, 60°C, or 70°C.

In other preferred embodiments, the DNA polymerase  
10 has less than 1000, 250, 100, 50, 10 or even 2 units of exonuclease activity per mg of polymerase (measured by standard procedure, see below) and is able to utilize primers having only 4, 6 or 10 bases; and the concentration of all four deoxynucleoside triphosphates  
15 at the start of the incubating step is sufficient to allow DNA synthesis to continue until terminated by the agent, e.g., a ddNTP.

For cycle sequencing, the DNA polymerases of the present invention make it possible to use significantly  
20 lower amounts of dideoxynucleotides compared to naturally occurring enzymes. That is, the method involves providing an excess amount of deoxynucleotides to all four dideoxynucleotides in a cycle sequencing reaction, and performing the cycle sequencing reaction.  
25 Preferably, more than 2, 5, 10 or even 100 fold excess of a dNTP is provided to the corresponding ddNTP.

In a related aspect, the invention features a kit or solution for DNA sequencing including a DNA polymerase of the present invention and a reagent  
30 necessary for the sequencing such as dITP, deaza GTP, a

chain terminating agent such as a ddNTP, and a manganese-containing solution or powder and optionally a pyrophosphatase.

In another aspect, the invention features a method  
5 for providing a DNA polymerase of the present invention by providing a nucleic acid sequence encoding the modified DNA polymerase, expressing the nucleic acid within a host cell, and purifying the DNA polymerase from the host cell.

10 In another related aspect, the invention features a method for sequencing a strand of DNA essentially as described above with one or more (preferably 2, 3 or 4) deoxyribonucleoside triphosphates, a DNA polymerase of the present invention, and a first chain terminating  
15 agent. The DNA polymerase causes the primer to be elongated to form a first series of first DNA products differing in the length of the elongated primer, each first DNA product having a chain terminating agent at its elongated end, and the number of molecules of each  
20 first DNA products being approximately the same for substantially all DNA products differing in length by no more than 20 bases. The method also features providing a second chain terminating agent in the hybridized mixture at a concentration different from the first  
25 chain terminating agent, wherein the DNA polymerase causes production of a second series of second DNA products differing in the length of the elongated primer, with each second DNA product having the second chain terminating agent at its elongated end. The  
30 number of molecules of each second DNA product is

approximately the same for substantially all second DNA products differing in length from each other by from 1 to 20 bases, and is distinctly different from the number of molecules of all the first DNA products having a length differing by no more than 20 bases from that of said second DNA products.

In preferred embodiments, three or four such chain terminating agents can be used to make different products and the sequence reaction is provided with a magnesium ion, or even a manganese or iron ion (e.g., at a concentration between 0.05 and 100 mM, preferably between 1 and 10 mM); and the DNA products are separated according to molecular weight in four or less lanes of a gel.

In another related aspect, the invention features a method for sequencing a nucleic acid by combining an oligonucleotide primer, a nucleic acid to be sequenced, between one and four deoxyribonucleoside triphosphates, a DNA polymerase of the present invention, and at least two chain terminating agents in different amounts, under conditions favoring extension of the oligonucleotide primer to form nucleic acid fragments complementary to the nucleic acid to be sequenced. For example, the chain terminating agent may be a dideoxynucleotide terminator for adenine, guanine, cytosine or thymine. The method further includes separating the nucleic acid fragments by size and determining the nucleic acid sequence. The agents are differentiated from each other by intensity of a label in the primer extension products.

While it is common to use gel electrophoresis to separate DNA products of a DNA sequencing reaction, those in the art will recognize that other methods may also be used. Thus, it is possible to detect each of  
5 the different fragments using procedures such as time of flight mass spectrometry, electron microscopy, and single molecule detection methods.

The invention also features an automated DNA sequencing apparatus having a reactor including reagents  
10 which provide at least two series of DNA products formed from a single primer and a DNA strand. Each DNA product of a series differs in molecular weight and has a chain terminating agent at one end. The reagents include a DNA polymerase of the present invention. The apparatus  
15 includes a separating means for separating the DNA product along one axis of the separator to form a series of bands. It also includes a band reading means for determining the position and intensity of each band after separation along the axis, and a computing means  
20 that determines the DNA sequence of the DNA strand solely from the position and intensity of the bands along the axis and not from the wavelength of emission of light from any label that may be present in the separating means.

25 Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

#### Description of the Preferred Embodiments

The drawings will first briefly be described.

### Drawings

Figs 1-4 are the DNA sequences, and corresponding amino acid sequences, of FY2, FY3, and the DNA polymerases of T. flavus and Thermus thermophilus, respectively. Figure 5 is the DNA sequence and corresponding amino acid sequence of FY4.

### Examples

The following examples serve to illustrate the DNA polymerases of the present invention and their use in sequencing.

#### Preparation of FY DNA Polymerases (FY2 and FY3)

##### Bacterial Strains

*E. coli* strains: MV1190 [ $\Delta$ (*srl* - *recA*) 306::Tn10,  $\Delta$ (*lac-proAB*), *thi*, *supE*, F' (*traD36 proAB<sup>+</sup> lacI<sup>q</sup> lacZ*  $\Delta$ M15)]; DH $\alpha$  [*gyrA96*, *recA1*, *relA1*, *endA1*, *thi-1*, *hsdR17*, *supE44*,  $\lambda^+$ ]; M5248 [ $\lambda$ (*bio275*, *ci857*, *cIII+*, *N+*,  $\Delta$  (*H1*))].

##### PCR

Reaction conditions based on the procedure of Barnes (91 Proc. Nat'l. Acad. Sci. 2216-2220, 1994) were as follows: 20mM Tricine pH8.8, 85mM KOAc, 200mM dNTPs, 10% glycerol, 5% DMSO, 0.5mM each primer, 1.5mM MgOAc, 2.5 U HotTub (Amersham Life Science Inc.), 0.025 U DeepVent (New England Biolabs), 1-100 ng target DNA per 100ml reaction. Cycling conditions were 94°C 30s, 68°C 10m40s for 8 cycles; then 94°C 30s, 68°C 12m00s for 8 cycles; then 94°C 30s, 68°C 13m20s for 8 cycles; then 94°C 30s, 68°C 14m40s for 8 cycles.

*In vitro* mutagenesis

Restriction enzyme digestions, plasmid preparations, and other *in vitro* manipulations of DNA were performed using standard protocols (Sambrook et al., Molecular Cloning 2nd Ed. Cold Spring Harbor Press, 1989). PCR (see protocol above) was used to introduce a Phe to Tyr amino acid change at codon 667 of native Taq DNA polymerase (which is codon 396 of FY2). Oligonucleotide primer 1 dGCTTGGGCAGAGGATCCGCCGGG (SEQ. ID. NO. 3) spans nucleotides 954 to 976 of the coding region of SEQ. ID. NO. 1 including a BamHI restriction site. Mutagenic oligo primer 2 dGGGATGGCTAGCTCCTGGGAGAGCGGTGGGCCGACATGCCGTAGA GGACCCCGTAGTTGATGG (SEQ. ID. NO. 4) spans nucleotides 1178 to 1241 including an NheI site and codon 396 of Sequence ID. NO. 1. A clone of *exo*-Taq deleted for the first 235 amino acids, pWB253 encoding DeltaTaq polymerase (Barnes, 112 *Gene* 29-35, 1992) was used as template DNA. Any clone of Taq polymerase or genomic DNA from *Thermus aquaticus* could also be utilized to amplify the identical PCR product. The PCR product was digested with BamHI and NheI, and this fragment was ligated to BamHI/NheI digested pWB253 plasmid to replace the corresponding fragment to create pWB253Y, encoding polymerase FY1. Cells of *E. coli* strain MV1190 were used for transformation and induction of protein expression, although any host strain carrying a *lac* repressor could be substituted. DNA sequencing verified the Phe to Tyr change in the coding region.

PCR primer 3 dGGAATTCATATGGACGATCTGAAGCTCTCC (SEQ. ID. NO. 5) spanning the start codon and containing restriction enzyme sites, was used with PCR primer 4 dGGGGTACCAAGCTTCACTCCTTGGCGGAGAG (SEQ. ID. NO. 6) containing restriction sites and spanning the stop codon (codon 562 of Sequence ID. NO. 1). A methionine start codon and restriction enzyme recognition sequences were added to PCR primer 5 dGGAATTCATATGCTGGAGAGGCTTGAGTT (SEQ. ID. NO. 7), which was used with primer 4 above.

PCR was performed using the above primer pairs, and plasmid pWB253Y as template. The PCR products were digested with restriction enzymes NdeI and KpnI and ligated to NdeI/KpnI digested vector pRE2 (Reddi et al., 17 Nucleic Acids Research 10,473-10,488, 1989) to make plasmids pRE236Y, encoding FY1 polymerase, and pRE273Y encoding FY2 polymerase, respectively. Cells of *E. coli* strain DHA<sup>+</sup> were used for primary transformation with this and all subsequent pRE2 constructions, and strain M5248 ( $\Delta$ cI857) was used for protein expression, although any comparable pair of *E. coli* strains carrying the cI<sup>+</sup> and cI857 alleles could be utilized. Alternatively, any rec<sup>+</sup> cI<sup>+</sup> strain could be induced by chemical agents such as nalidixic acid to produce the polymerase. The sequences of both genes were verified. pRE273Y was found to produce a single polypeptide band on SDS polyacrylamide gels, unlike pRE253Y or pRE236Y.

Primer 6 dGGAATTCATATGCTGGAACGCTGGAGTTGGCAGCCTC CTC (SEQ. ID. NO. 8) and primer 4 were used to make a PCR product introducing silent changes in codon usage of FY2. The product was digested with NdeI/BamHI and



15

ligated to a pRE2 construct containing the 3' end of FY2 to create pREFY2pref, encoding FY2 DNA polymerase.

Primer 7 dGGAATTCATATGGCTCTGGAACGTCTGGAGTTTGGCAGCCTCCTC (SEQ. ID. NO. 9) and primer 4 were used to make a PCR

- 5 product introducing an additional alanine codon commonly occurring at the second position of highly expressed genes. The NdeI/BamHI digested fragment was used as above to create pREFY3, encoding FY3 DNA polymerase.

#### Preparation of FY4 DNA Polymerase

##### 10 Bacterial Strains

*E. coli* strains: DH1 $\lambda^+$  [*gyrA96*, *recA1*, *relA1*, *endA1*, *thi-1*, *hsdR17*, *supE44*,  $\lambda^+$ ]; M5248 [ $\lambda$  (*bio275*, *cI857*, *cIII+*, *N+*,  $\Delta$  (*H1*))].

##### PCR

- 15 Genomic DNA was prepared by standard techniques from *Thermus thermophilus*. The DNA polymerase gene of *Thermus thermophilus* is known to reside on a 3 kilobase AlwNI fragment. To enrich for polymerase sequences in some PCR reactions, the genomic DNA was digested prior
- 20 to PCR with AlwNI, and fragments of approximately 3 kb were selected by agarose gel electrophoresis to be used as template DNA. Reaction conditions were as follows: 10mM Tris pH8.3, 50mM KCl, 800 $\mu$ M dNTPs, 0.001% gelatin, 1.0 $\mu$ M each primer, 1.5mM MgCl<sub>2</sub>, 2.5 U Tth, 0.025 U
- 25 DeepVent (New England Biolabs), per 100 $\mu$ l reaction. Cycling conditions were 94°C 2 min, then 35 cycles of 94°C 30s, 55°C 30s, 72°C 3 min, followed by 72 °C for 7 min.

### *In vitro* mutagenesis

- Restriction enzyme digestions, plasmid preparations, and other *in vitro* manipulations of DNA were performed using standard protocols (Sambrook et al., 1989). Plasmid pMR1 was constructed to encode an exonuclease-free polymerase, with optimized codons for expression in *E. coli* at the 5' end. Primer 8 (SEQ. ID. NO. 10) (GGAATTCCATATGCTGGAACGTCTGGAATTCGGCAGCCTC) was used with Primer 9 (SEQ. ID. NO. 11) (GGGGTACCCTAACCCTTGGCGGAAAGCCAGTC) to create a PCR product from *Tth* genomic DNA, which was digested with restriction enzymes NdeI and KpnI and inserted into plasmid pRE2 (Reddi et al., 1989, Nucleic Acids Research 17, 10473 - 10488) digested with the same enzymes.
- To create the desired F396Y mutation, two PCR products were made from *Tth* chromosomal DNA. Primer 8 above was used in combination with Primer 10 (SEQ. ID. NO. 12) (GGGATGGCTAGCTCCTGGGAGAGCCTATGGGCGGACATGCCGTAGAGGACGCCGTAGTTCACCG) to create a portion of the gene containing the F to Y amino acid change as well as a silent change to create an NheI restriction site. Primer 11 (SEQ. ID. NO. 13) (CTAGCTAGCCATCCCCCTACGAAGAAGCGGTGGCCT) was used in combination with primer 9 above to create a portion of the gene from the introduced NheI site to the stop codon at the 3' end of the coding sequence. The PCR product of Primers 8 and 10 was digested with NdeI and NheI, and the PCR product of Primers 9 and 11 was digested with NheI and KpnI. These were introduced into expression vector pRE2 which was digested with NdeI and KpnI to produce plasmid pMR5.

In addition to the desired changes, pMR5 was found to have a spurious change introduced by PCR, which led to an amino acid substitution, K234R. Plasmid pMR8 was created to eliminate this substitution, by replacing the  
5 AflIII/BamHI fragment of pMR5 for the corresponding fragment from pMR1. The FY4 polymerase encoded by plasmid pMR8 (SEQ. ID. NO. 14) is given in Figure 5.

Cells of *E. coli* strain DH1 $\lambda^+$  were used for primary transformation, and strain M5248 ( $\lambda$ cI857) was used for  
10 protein expression, although any comparable pair of *E. coli* strains carrying the cI $^+$  and cI857 alleles could be utilized. Alternatively, any rec $^+$  cI $^+$  strain could be induced by chemical agents such as nalidixic acid to produce the polymerase.

#### 15 Protein Sequencing

Determinations of amino terminal protein sequences were performed at the W.M. Keck Foundation, Biotechnology Resource Laboratory, New Haven, Connecticut.

#### 20 Purification of Polymerases

A 1 liter culture of 2X LB (2% Bacto-Tryptone, 1% Bacto-Yeast Extract, 0.5% NaCl) + 0.2% Casamino Acids + 20 mM KPO $_4$  pH 7.5 + 50  $\mu$ g/ml Ampicillin was inoculated with a glycerol stock of the appropriate cell strain and  
25 grown at 30°C with agitation until cells were in log phase (0.7-1.0 OD $_{590}$ ). 9 liters of 2X LB + 0.2% Casamino Acids + 20 mM KPO $_4$  pH 7.5 + 0.05% Mazu Anti-foam was inoculated with 1 liter of log phase cells in 10 liter Microferm Fermentors (New Brunswick Scientific Co.).  
30 Cells were grown at 30°C under 15 psi pressure, 350-450

rpm agitation, and an air flow rate of 14,000 cc/min  $\pm$ 1000 cc/min. When the OD<sub>590</sub> reached 1.5-2.0, the cultures were induced by increasing the temperature to 40-42°C for 90-120 minutes. The cultures were then

- 5 cooled to < 20°C and the cells harvested by centrifugation in a Sorvall RC-3B centrifuge at 5000 rpm at 4°C for 15-20 minutes. Harvested cells were stored at -80°C.

- Frozen cells were broken into small pieces and
- 10 resuspended in pre-warmed (90-95°C) Lysis Buffer (20 mM Tris pH 8.5, 1 mM EDTA, 10 mM MgCl<sub>2</sub>, 16 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.1% Tween 20, 0.1% Nonidet P-40, 1 mM PMSF). Resuspended cells were then heated rapidly to 80°C and incubated at 80°C for 20 minutes with constant stirring. The
- 15 suspension was then rapidly cooled on ice. The cell debris was removed by centrifugation using a Sorvall GSA rotor at 10,000 rpm for 20 minutes at 4°C. The NaCl concentration of the supernatant was adjusted to 300 mM. The sample was then passed through a diethylaminoethyl
- 20 cellulose (Whatman DE-52) column that had been previously equilibrated with Buffer A (20 mM Tris pH 8.5, 1 mM EDTA, 0.1% Tween 20, 0.1% Nonidet P-40, 300 mM NaCl, 10% glycerol, 1 mM DTT), and polymerase collected in the flow through. The sample was then diluted to a
- 25 concentration of NaCl of 100mM and applied to a Heparin-sepharose column. The polymerase was eluted from the column with a NaCl gradient (100-500 mM NaCl). The sample was then dialyzed against Buffer B (20 mM Tris pH 8.5, 1 mM EDTA, 0.1% Tween 20, 0.1% Nonidet P-40, 10 mM
- 30 KCl, 10% glycerol, 1 mM DTT) and further diluted as

needed to lower the conductivity of the sample to the conductivity of Buffer B. The sample was then applied to a diethylaminoethyl (Waters DEAE 15 HR) column and eluted with a 10-500 mM KCl gradient. The polymerase  
5 was then diluted with an equal volume of Final Buffer (20 mM Tris pH 8.5, 0.1 mM EDTA, 0.5% Tween 20, 0.5% Nonidet P-40, 100 mM KCl, 50% glycerol, 1 mM DTT) and dialyzed against Final Buffer.

#### Assay of Exonuclease Activity

10 The exonuclease assay was performed by incubating 5 ul (25-150 units) of DNA polymerase with 5 ug of labelled [<sup>3</sup>H]-pBR322 PCR fragment (1.6x10<sup>4</sup> cpm/ug DNA) in 100 ul of reaction buffer of 20 mM Tris-HCl pH 8.5, 5 mM MgCl<sub>2</sub>, 10 mM KCl, for 1 hour at 60 °C. After this time  
15 interval, 200 ul of 1:1 ratio of 50 ug/ml salmon sperm DNA with 2 mM EDTA and 20% TCA with 2% sodium pyrophosphate were added into the assay aliquots. The aliquots were put on ice for 10 min and then centrifuged at 12,000g for 10 min. Acid-soluble radioactivity in 200  
20 ul of the supernatant was quantitated by liquid scintillation counting. One unit of exonuclease activity was defined as the amount of enzyme that catalyzed the acid solubilization of 10 nmol of total nucleotide in 30 min at 60 °C.

#### 25 Utility in DNA Sequencing

Example 1: DNA Sequencing with FY Polymerases (e.g., FY2 and FY3)

The following components were added to a microcentrifuge vial (0.5 ml) : 0.4 pmol M13 DNA (e.g., M13mp18, 1.0  $\mu$ g); 2  $\mu$ l Reaction Buffer ( 260 mM Tris-HCl, pH 9.5 65 mM  $MgCl_2$ ); 2  $\mu$ l of labeling nucleotide mixture (1.5  $\mu$ M each of dGTP, dCTP and dTTP); 0.5  $\mu$ l (5  $\mu$ Ci) of [ $\alpha$ - $^{32}P$ ]dATP (about 2000Ci/mmol); 1  $\mu$ l -40 primer (0.5  $\mu$ M; 0.5 pmol/ $\mu$ l 5'GTTTCCCAGTCACGAC-3'); 2  $\mu$ l of a mixture containing 4 U/ $\mu$ l FY polymerase and 6.6 U/ml *Thermoplasma acidophilum* inorganic pyrophosphatase (32 U/ $\mu$ l polymerase and 53 U/ml pyrophosphatase in 20 mM Tris (pH8.5), 100 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5% NP-40, 0.5% TWEEN-20 and 50% glycerol, diluted 8 fold in dilution buffer (10 mM Tris-HCl pH8.0, 1 mM 2-mercaptoethanol, 0.5% TWEEN-20, 0.5% NP-40)); and water to a total volume of 17.5  $\mu$ l. These components (the labeling reaction) were mixed and the vial was placed in a constant-temperature water bath at 45°C for 5 minutes.

Four vials were labeled A, C, G, and T, and filled with 4  $\mu$ l of the corresponding termination mix: dda termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddATP); ddT termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddTTP); ddC termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddCCTP); ddG termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddGTP).

The labeling reaction was divided equally among the four termination vials (4  $\mu$ l to each termination reaction vial), and tightly capped.

The four vials were placed in a constant-temperature water bath at 72°C for 5 minutes. Then 4  $\mu$ l

- of Stop Solution (95% Formamide 20 mM EDTA, 0.05% Bromophenol Blue, 0.05% Xylene Cyanol FF) added to each vial, and heated briefly to 70°-80°C immediately prior to loading on a sequencing gel (8% acrylamide, 8.3 M urea).
- 5 Autoradiograms required an 18-36 hour exposure using Kodak XAR-5 film or Amersham Hyperfilm MP. High-quality sequence results with uniform band intensities were obtained. The band intensities were much more uniform than those obtained with similar protocols using *Taq* DNA
- 10 polymerase or  $\Delta$ *Taq* DNA polymerase.

Example 2: DNA Cycle Sequencing with FY Polymerases

- The following components were added to a microcentrifuge vial (0.5 ml) which which is suitable for insertion into a thermocycler machine (e.g., Perkin-
- 15 Elmer DNA Thermal Cycler): 0.05 pmol or more M13 DNA (e.g., M13mp18, 0.1  $\mu$ g), or 0.1  $\mu$ g double-stranded plasmid DNA (e.g., pUC19); 2  $\mu$ l Reaction Buffer (260 mM Tris-HCl, pH 9.5 65 mM MgCl<sub>2</sub>); 1  $\mu$ l 3.0  $\mu$ M dGTP; 1  $\mu$ l 3.0  $\mu$ M dTTP; 0.5  $\mu$ l (5  $\mu$ Ci) of [ $\alpha$ -<sup>32</sup>P]dATP (about
- 20 2000Ci/mmol); 1  $\mu$ l -40 primer (0.5  $\mu$ M; 0.5 pmol/ $\mu$ l 5'-GTTTTCACGTCACGAC-3'); 2  $\mu$ l of a mixture containing 4 U/ $\mu$ l FY polymerase and 6.6 U/ml *Thermoplasma acidophilum* inorganic pyrophosphatase (32 U/ $\mu$ l polymerase and 53 U/ml pyrophosphatase in 20 mM Tris (pH8.5), 100
- 25 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5% NP-40, 0.5% TWEEN-20 and 50% glycerol, diluted 8 fold in dilution buffer (10 mM Tris-HCl pH8.0, 1 mM 2-mercaptoethanol, 0.5% TWEEN-20, 0.5% NP-40)); and water to a total volume of 17.5  $\mu$ l.

These components (labeling reaction mixture) were mixed and overlaid with 10  $\mu$ l light mineral oil (Amersham). The vial was placed in the thermocycler and 30-100 cycles (more than 60 cycles is unnecessary) from 45°C for 1 minute to 95°C for 0.5 minute performed. (Temperatures can be cycled from 55°-95°C, if desired). The temperatures may be adjusted if the melting temperature of the primer/template is significantly higher or lower, but these temperatures work well for most primer-templates combinations. This step can be completed in about 3 minutes per cycle.

Four vials were labeled A, C, G, and T, and filled with 4 ml of the corresponding termination mix: ddA termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddATP); ddT termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddTTP); ddC termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddCTP); ddG termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddGTP). No additional enzyme is added to the termination vials. The enzyme carried in from the prior (labeling) step is sufficient.

The cycled labeling reaction mixture was divided equally among the four termination vials (4  $\mu$ l to each termination reaction vial), and overlaid with 10  $\mu$ l of light mineral oil.

The four vials were placed in the thermocycler and 30-200 cycles (more than 60 cycles is unnecessary) performed from 95°C for 15 seconds, 55°C for 30 seconds, and 72°C for 120 seconds. This step was conveniently



completed overnight. Other times and temperatures are also effective.

Six  $\mu$ l of reaction mixture was removed (avoiding oil), 3  $\mu$ l of Stop Solution (95% Formamide 20 mM EDTA, 0.05% Bromophenol Blue, 0.05% Xylene Cyanol FF) added, and heated briefly to 70°-80°C immediately prior to loading on a sequencing gel. Autoradiograms required an 18-36 hour exposure using Kodak XAR-5 film or Amersham Hyperfilm MP. High-quality sequence results with uniform band intensities were obtained. The band intensities were much more uniform than those obtained with similar protocols using Taq DNA polymerase or  $\Delta$ Taq DNA polymerase.

Example 3: Sequencing with dGTP analogs to eliminate compression artifacts.

For either of the sequencing methods outlined in examples 1 and 2, 7-Deaza-2'deoxy-GTP can be substituted for dGTP in the labeling and termination mixtures at exactly the same concentration as dGTP. When this substitution is made, secondary structures on the gels are greatly reduced. Similarly, 2'-deoxyinosinetriphosphate can also be substituted for dGTP but its concentration must be 10-fold higher than the corresponding concentration of dGTP. Substitution of dITP for dGTP is even more effective in eliminating compression artifacts than 7-deaza-dGTP.

Example 4: Other Sequencing methods using FY polymerases

FY polymerases have been adapted for use with many other sequencing methods, including the use of  
5 fluorescent primers and fluorescent-dideoxy-terminators for sequencing with the ABI 373A DNA sequencing instrument.

Example 5: SDS-Polyacrylamide Gel Electrophoresis

Protein samples were run on a 14 X 16 mm 7.5 or 10%  
10 polyacrylamide gel. (Gels were predominantly 10% Polyacrylamide using a 14 X 16 mm Hoefer apparatus. Other sizes, apparatuses, and percentage gels are acceptable. Similar results can also be obtained using the Pharmacia Phast Gel system with SDS, 8-25% gradient  
15 gels. Reagent grade and ultrapure grade reagents were used.) The stacking gel consisted of 4% acrylamide (30:0.8, acrylamide: bisacrylamide), 125 mM Tris-HCl pH 6.8, 0.1% Sodium Dodecyl Sulfate (SDS). The resolving gel consisted of 7.5 or 10% acrylamide (30:0.8,  
20 acrylamide: bisacrylamide), 375 mM Tris-HCl pH 8.8, 0.1% SDS. Running Buffer consisted of 25 mM Tris, 192 mM Glycine and 0.1% SDS. 1X Sample Buffer consisted of 25 mM Tris-HCl pH 6.8, 0.25% SDS, 10% Glycerol, 0.1M Dithiothreitol, 0.1% Bromophenol Blue, and 1mM EDTA. A  
25 1/4 volume of 5X Sample Buffer was added to each sample. Samples were heated in sample buffer to 90-100°C for approximately 5 minutes prior to loading. A 1.5 mm thick gel was run at 50-100 mA constant current for 1-3 hours (until bromophenol blue was close to the bottom of

the gel). The gel was stained with 0.025% Coomassie  
Blue R250 in 50% methanol, 10% acetic acid and destained  
in 5% methanol, 7% acetic acid solution. A record of  
the gel was made by taking a photograph of the gel, by  
5 drying the gel between cellulose film sheets, or by  
drying the gel onto filter paper under a vacuum.

Other embodiments are within the following claims.

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- (i) APPLICANT: AMERSHAM LIFE SCIENCE
- 5 (ii) TITLE OF INVENTION: THERMOSTABLE DNA  
POLYMERASES
- (iii) NUMBER OF SEQUENCES: 14
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- 10 (A) ADDRESSEE: Lyon & Lyon  
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(E) COUNTRY: U.S.A.  
(F) ZIP: 90071-2066
- 15 (v) COMPUTER READABLE FORM:
- 20 (A) MEDIUM TYPE: 3.5" Diskette, 1.44 Mb  
storage  
(B) COMPUTER: IBM Compatible  
(C) OPERATING SYSTEM: IBM P.C. DOS 5.0  
(D) SOFTWARE: Word Perfect 5.1
- (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER: To Be Assigned  
(B) FILING DATE:  
(C) CLASSIFICATION:
- 25 (vii) PRIOR APPLICATION DATA:

27

Prior applications total,  
including application  
described below: one

- 5 (A) APPLICATION NUMBER: US 08/455,686  
(B) FILING DATE: May 31, 1995

(viii) ATTORNEY/AGENT INFORMATION:

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(B) REGISTRATION NUMBER: 32,327  
(C) REFERENCE/DOCKET NUMBER: 219/304-PCT

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(2) INFORMATION FOR SEQ ID NO: 1:

15 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1686 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

20 (ix) FEATURE:

- (A) NAME/KEY: FY2  
(B) LOCATION: 1...1683

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

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1 5 10 15	
CTG GAA AGC CCC AAG GCC CTG GAG GAG GCC CCC TGG CCC CCG CCG GAA	96
Leu Glu Ser Pro Lys Ala Leu Glu Glu Ala Pro Trp Pro Pro Pro Glu	
20 25 30	
30 GGG GCC TTC GTG GGC TTT GTG CTT TCC CGC AAG GAG CCC ATG TGG GCC	144

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	Gly	Ala	Phe	Val	Gly	Phe	Val	Leu	Ser	Arg	Lys	Glu	Pro	Met	Trp	Ala	
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	CCC	GAG	CCT	TAT	AAA	GCC	CTC	AGG	GAC	CTG	AAG	GAG	GGC	CGG	GGG	CTT	240
	Pro	Glu	Pro	Tyr	Lys	Ala	Leu	Arg	Asp	Leu	Lys	Glu	Ala	Arg	Gly	Leu	
	65				70					75					80		
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	CCG	CCC	GGC	GAC	GAC	CCC	ATG	CTC	CTC	GCC	TAC	CTC	CTG	GAC	CCT	TCC	336
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	Leu	Trp	Gly	Arg	Leu	Glu	Gly	Glu	Glu	Arg	Leu	Leu	Trp	Leu	Tyr	Arg	
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25	Glu	Val	Glu	Arg	Pro	Leu	Ser	Ala	Val	Leu	Ala	His	Met	Glu	Ala	Thr	
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30	GCC	GAG	GAG	ATC	GCC	CGC	CTC	GAG	GCC	GAG	GTC	TTC	CGC	CTG	GCC	GGC	624
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	CAC	CCC	TTC	AAC	CTC	AAC	TCC	CGG	GAC	CAG	CTG	GAA	AGG	GTC	CTC	TTT	672
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				210				215						220			
	GAC	GAG	CTA	GGG	CTT	CCC	GCC	ATC	GGC	AAG	ACG	GAG	AAG	ACC	GGC	AAG	720
	Asp	Glu	Leu	Gly	Leu	Pro	Ala	Ile	Gly	Lys	Thr	Glu	Lys	Thr	Gly	Lys	
	225					230					235					240	

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5	ATC GTG GAG AAG ATC CTG CAG TAC CGG GAG CTC ACC AAG CTG AAG AGC	816
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	ACC TAC ATT GAC CCC TTG CCG GAC CTC ATC CAC CCC AGG ACG GGC CGC	864
	Thr Tyr Ile Asp Pro Leu Pro Asp Leu Ile His Pro Arg Thr Gly Arg	
	275 280 285	
10	CTC CAC ACC CGC TTC AAC CAG ACG GCC ACG GCC ACG GGC AGG CTA AGT	912
	Leu His Thr Arg Phe Asn Gln Thr Ala Thr Ala Thr Gly Arg Leu Ser	
	290 295 300	
	AGC TCC GAT CCC AAC CTC CAG AAC ATC CCC GTC CGC ACC CCG CTT GGG	960
	Ser Ser Asp Pro Asn Leu Gln Asn Ile Pro Val Arg Thr Pro Leu Gly	
15	305 310 315 320	
	CAG AGG ATC CGC CGG GCC TTC ATC GCC GAG GAG GGG TGG CTA TTG GTG	1008
	Gln Arg Ile Arg Arg Ala Phe Ile Ala Glu Glu Gly Trp Leu Leu Val	
	325 330 335	
20	GCC CTG GAC TAT AGC CAG ATA GAG CTC AGG GTG CTG GCC CAC CTC TCC	1056
	Ala Leu Asp Tyr Ser Gln Ile Glu Leu Arg Val Leu Ala His Leu Ser	
	340 345 350	
	GGC GAC GAG AAC CTG ATC CGG GTC TTC CAG GAG GGG CGG GAC ATC CAC	1104
	Gly Asp Glu Asn Leu Ile Arg Val Phe Gln Glu Gly Arg Asp Ile His	
	355 360 365	
25	ACG GAG ACC GCC AGC TGG ATG TTC GGC GTC CCC CGG GAG GCC GTG GAC	1152
	Thr Glu Thr Ala Ser Trp Met Phe Gly Val Pro Arg Glu Ala Val Asp	
	370 375 380	
	CCC CTG ATG CGC CGG GCG GCC AAG ACC ATC AAC TAC GGG GTC CTC TAC	1200
	Pro Leu Met Arg Arg Ala Ala Lys Thr Ile Asn Tyr Gly Val Leu Tyr	
30	385 390 395 400	
	GGC ATG TCG GCC CAC CGC CTC TCC CAG GAG CTA GCC ATC CCT TAC GAG	1248
	Gly Met Ser Ala His Arg Leu Ser Gln Glu Leu Ala Ile Pro Tyr Glu	
	405 410 415	
35	GAG GCC CAG GCC TTC ATT GAG CGC TAC TTT CAG AGC TTC CCC AAG GTG	1296
	Glu Ala Gln Ala Phe Ile Glu Arg Tyr Phe Gln Ser Phe Pro Lys Val	
	420 425 430	
	CGG GCC TGG ATT GAG AAG ACC CTG GAG GAG GGC AGG AGG CGG GGG TAC	1344
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	435 440 445	

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      GTG GAG ACC CTC TTC GGC CGC CGC CGC TAC GTG CCA GAC CTA GAG GCC 1392
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      CGG GTG AAG AGC GTG CGG GAG GCG GCC GAG CGC ATG GCC TTC AAC ATG 1440
      Arg Val Lys Ser Val Arg Glu Ala Ala Glu Arg Met Ala Phe Asn Met
      465                               470                               475                               480

      CCC GTC CAG GGC ACC GCC GCC GAC CTC ATG AAG CTG GCT ATG GTG AAG 1488
      Pro Val Gln Gly Thr Ala Ala Asp Leu Met Lys Leu Ala Met Val Lys
      485                               490                               495

10    CTC TTC CCC AGG CTG GAG GAA ATG GGG GCC AGG ATG CTC CTT CAG GTC 1536
      Leu Phe Pro Arg Leu Glu Glu Met Gly Ala Arg Met Leu Leu Gln Val
      500                               505                               510

      CAC GAC GAG CTG GTC CTC GAG GCC CCA AAA GAG AGG GCG GAG GCC GTG 1584
      His Asp Glu Leu Val Leu Glu Ala Pro Lys Glu Arg Ala Glu Ala Val
      515                               520                               525

      GCC CGG CTG GCC AAG GAG GTC ATG GAG GGG GTG TAT CCC CTG GCC GTG 1632
      Ala Arg Leu Ala Lys Glu Val Met Glu Gly Val Tyr Pro Leu Ala Val
      530                               535                               540

      CCC CTG GAG GTG GAG GTG GGG ATA GGG GAG GAC TGG CTC TCC GCC AAG 1680
      Pro Leu Glu Val Glu Val Gly Ile Gly Glu Asp Trp Leu Ser Ala Lys
      545                               550                               555                               560

      GAG TGA
      Glu *
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(2) INFORMATION FOR SEQ ID NO: 2:

25 (i) SEQUENCE CHARACTERISTICS:

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      (A) LENGTH:          1689 base pairs
      (B) TYPE:            nucleic acid
      (C) STRANDEDNESS:    single
      (D) TOPOLOGY:        linear

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30 (ix) FEATURE:

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      (A) NAME/KEY: FY3
      (B) LOCATION: 1...1686

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

ATG GCT CTG GAA CGT CTG GAG TTT GGC AGC CTC CTC CAC GAG TTC GGC 48



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				85					90					95			
	CTC	CCG	CCC	GGC	GAC	GAC	CCC	ATG	CTC	CTC	GCC	TAC	CTC	CTG	GAC	CCT	336
20	Leu	Pro	Pro	Gly	Asp	Asp	Pro	Met	Leu	Leu	Ala	Tyr	Leu	Leu	Asp	Pro	
				100				105					110				
	TCC	AAC	ACC	ACC	CCC	GAG	GGG	GTG	GCC	CGG	CGC	TAC	GGC	GGG	GAG	TGG	384
	Ser	Asn	Thr	Thr	Pro	Glu	Gly	Val	Ala	Arg	Arg	Tyr	Gly	Gly	Glu	Trp	
			115					120					125				
	ACG	GAG	GAG	GCG	GGG	GAG	CGG	GCC	GCC	CTT	TCC	GAG	AGG	CTC	TTC	GCC	432
25	Thr	Glu	Glu	Ala	Gly	Glu	Arg	Ala	Ala	Leu	Ser	Glu	Arg	Leu	Phe	Ala	
		130					135					140					
	AAC	CTG	TGG	GGG	AGG	CTT	GAG	GGG	GAG	GAG	AGG	CTC	CTT	TGG	CTT	TAC	480
	Asn	Leu	Trp	Gly	Arg	Leu	Glu	Gly	Glu	Glu	Arg	Leu	Leu	Trp	Leu	Tyr	
	145				150					155					160		
30	CGG	GAG	GTG	GAG	AGG	CCC	CTT	TCC	GCT	GTC	CTG	GCC	CAC	ATG	GAG	GCC	528
	Arg	Glu	Val	Glu	Arg	Pro	Leu	Ser	Ala	Val	Leu	Ala	His	Met	Glu	Ala	
				165						170				175			
	ACG	GGG	GTG	CGC	CTG	GAC	GTG	GCC	TAT	CTC	AGG	GCC	TTG	TCC	CTG	GAG	576
35	Thr	Gly	Val	Arg	Leu	Asp	Val	Ala	Tyr	Leu	Arg	Ala	Leu	Ser	Leu	Glu	
			180					185					190				
	GTG	GCC	GAG	GAG	ATC	GCC	CGC	CTC	GAG	GCC	GAG	GTC	TTC	CGC	CTG	GCC	624
	Val	Ala	Glu	Glu	Ile	Ala	Arg	Leu	Glu	Ala	Glu	Val	Phe	Arg	Leu	Ala	
		195					200						205				
	GGC	CAC	CCC	TTC	AAC	CTC	AAC	TCC	CGG	GAC	CAG	CTG	GAA	AGG	GTC	CTC	672

32

	Gly	His	Pro	Phe	Asn	Leu	Asn	Ser	Arg	Asp	Gln	Leu	Glu	Arg	Val	Leu	
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5	Phe	Asp	Glu	Leu	Gly	Leu	Pro	Ala	Ile	Gly	Lys	Thr	Glu	Lys	Thr	Gly	
	225					230					235				240		
	AAG	CGC	TCC	ACC	AGC	GCC	GCC	GTC	CTG	GAG	GCC	CTC	CGC	GAG	GCC	CAC	768
	Lys	Arg	Ser	Thr	Ser	Ala	Ala	Val	Leu	Glu	Ala	Leu	Arg	Glu	Ala	His	
					245				250					255			
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10	Pro	Ile	Val	Glu	Lys	Ile	Leu	Gln	Tyr	Arg	Glu	Leu	Thr	Lys	Leu	Lys	
				260					265					270			
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	Ser	Thr	Tyr	Ile	Asp	Pro	Leu	Pro	Asp	Leu	Ile	His	Pro	Arg	Thr	Gly	
				275					280					285			
15	CGC	CTC	CAC	ACC	CGC	TTC	AAC	CAG	ACG	GCC	ACG	GCC	ACG	GGC	AGG	CTA	912
	Arg	Leu	His	Thr	Arg	Phe	Asn	Gln	Thr	Ala	Thr	Ala	Thr	Gly	Arg	Leu	
		290					295						300				
	AGT	AGC	TCC	GAT	CCC	AAC	CTC	CAG	AAC	ATC	CCC	GTC	CGC	ACC	CCG	CTT	960
20	Ser	Ser	Ser	Asp	Pro	Asn	Leu	Gln	Asn	Ile	Pro	Val	Arg	Thr	Pro	Leu	
	305					310					315				320		
	GGG	CAG	AGG	ATC	CGC	CGG	GCC	TTC	ATC	GCC	GAG	GAG	GGG	TGG	CTA	TTG	1008
	Gly	Gln	Arg	Ile	Arg	Arg	Ala	Phe	Ile	Ala	Glu	Glu	Gly	Trp	Leu	Leu	
				325					330					335			
	GTG	GCC	CTG	GAC	TAT	AGC	CAG	ATA	GAG	CTC	AGG	GTG	CTG	GCC	CAC	CTC	1056
25	Val	Ala	Leu	Asp	Tyr	Ser	Gln	Ile	Glu	Leu	Arg	Val	Leu	Ala	His	Leu	
				340					345					350			
	TCC	GGC	GAC	GAG	AAC	CTG	ATC	CGG	GTC	TTC	CAG	GAG	GGG	CGG	GAC	ATC	1104
	Ser	Gly	Asp	Glu	Asn	Leu	Ile	Arg	Val	Phe	Gln	Glu	Gly	Arg	Asp	Ile	
				355					360					365			
30	CAC	ACG	GAG	ACC	GCC	AGC	TGG	ATG	TTC	GGC	GTC	CCC	CGG	GAG	GCC	GTG	1152
	His	Thr	Glu	Thr	Ala	Ser	Trp	Met	Phe	Gly	Val	Pro	Arg	Glu	Ala	Val	
				370					375					380			
	GAC	CCC	CTG	ATG	CGC	CGG	GCG	GCC	AAG	ACC	ATC	AAC	TAC	GGG	GTC	CTC	1200
35	Asp	Pro	Leu	Met	Arg	Arg	Ala	Ala	Lys	Thr	Ile	Asn	Tyr	Gly	Val	Leu	
				385			390				395				400		
	TAC	GGC	ATG	TCG	GCC	CAC	CGC	CTC	TCC	CAG	GAG	CTA	GCC	ATC	CCT	TAC	1248
	Tyr	Gly	Met	Ser	Ala	His	Arg	Leu	Ser	Gln	Glu	Leu	Ala	Ile	Pro	Tyr	
					405					410					415		

33

GAG GAG GCC CAG GCC TTC ATT GAG CGC TAC TTT CAG AGC TTC CCC AAG 1296  
 Glu Glu Ala Gln Ala Phe Ile Glu Arg Tyr Phe Gln Ser Phe Pro Lys  
 420 425 430

5 GTG CGG GCC TGG ATT GAG AAG ACC CTG GAG GAG GGC AGG AGG CGG GGG 1344  
 Val Arg Ala Trp Ile Glu Lys Thr Leu Glu Glu Gly Arg Arg Arg Gly  
 435 440 445

TAC GTG GAG ACC CTC TTC GGC CGC CGC CGC TAC GTG CCA GAC CTA GAG 1392  
 Tyr Val Glu Thr Leu Phe Gly Arg Arg Arg Tyr Val Pro Asp Leu Glu  
 450 455 460

10 GCC CGG GTG AAG AGC GTG CGG GAG GCG GCC GAG CGC ATG GCC TTC AAC 1440  
 Ala Arg Val Lys Ser Val Arg Glu Ala Ala Glu Arg Met Ala Phe Asn  
 465 470 475 480

15 ATG CCC GTC CAG GGC ACC GCC GCC GAC CTC ATG AAG CTG GCT ATG GTG 1488  
 Met Pro Val Gln Gly Thr Ala Ala Asp Leu Met Lys Leu Ala Met Val  
 485 490 495

AAG CTC TTC CCC AGG CTG GAG GAA ATG GGG GCC AGG ATG CTC CTT CAG 1536  
 Lys Leu Phe Pro Arg Leu Glu Glu Met Gly Ala Arg Met Leu Leu Gln  
 500 505 510

20 GTC CAC GAC GAG CTG GTC CTC GAG GCC CCA AAA GAG AGG GCG GAG GCC 1584  
 Val His Asp Glu Leu Val Leu Glu Ala Pro Lys Glu Arg Ala Glu Ala  
 515 520 525

GTG GCC CGG CTG GCC AAG GAG GTC ATG GAG GGG GTG TAT CCC CTG GCC 1632  
 Val Ala Arg Leu Ala Lys Glu Val Met Glu Gly Val Tyr Pro Leu Ala  
 530 535 540

25 GTG CCC CTG GAG GTG GAG GTG GGG ATA GGG GAG GAC TGG CTC TCC GCC 1680  
 Val Pro Leu Glu Val Glu Val Gly Ile Gly Glu Asp Trp Leu Ser Ala  
 545 550 555 560

AAG GAG TGA 1689  
 Lys Glu \*

30 (2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 35 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

34

GCTTGGGCAG AGGATCCGCC GGG

23

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 64 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

GGGATGGCTA GCTCCTGGGA GAGGCGGTGG GCCGACATGC CGTAGAGGAC 50  
10 CCCGTAGTTG ATGG 64

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

GGAATTCCAT ATGGACGATC TGAAGCTCTC C 31

(2) INFORMATION FOR SEQ ID NO: 6:

20 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

35

GGGGTACCAA GCTTCACTCC TTGGCGGAGA G

31

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

GGAATTCAT ATGCTGGAGA GGCTTGAGTT T

31

10 (2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 43 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

GGAATTCAT ATGCTGGAAC GTCTGGAGTT TGGCAGCCTC CTC

43

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

20 (A) LENGTH: 46 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

36

GGAATTCCAT ATGGCTCTGG AACGTCTGGA GTTGGCAGC CTCCTC

46

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 40 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

GGAATTCCAT ATGCTGGAAC GTCTGGAATT CGGCAGCCTC

40

10 (2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 32 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

GGGGTACCCT AACCTTGGC GGAAAGCCAG TC

32

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

20 (A) LENGTH: 64 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

25 GGGATGGCTA GCTCCTGGGA GAGCCTATGG GCGGACATGC CGTAGAGGAC

50

37

GCCGTAGTTC ACCG

64

(2) INFORMATION FOR SEQ ID NO: 13:

## (i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 35 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

CTAGCTAGCC ATCCCCTACG AAGAAGCGGT GGCCT

35

10 (2) INFORMATION FOR SEQ ID NO: 14:

## (i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 1686 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## (ix) FEATURE:

(A) NAME/KEY: FY4  
 (B) LOCATION: 1...1683

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

20 ATG CTG GAA CGT CTG GAA TTC GGC AGC CTC CTC CAC GAG TTC GGC CTC 48  
 Met Leu Glu Arg Leu Glu Phe Gly Ser Leu Leu His Glu Phe Gly Leu  
 1 5 10 15

CTG GAG GCC CCC GCC CCC CTG GAG GAG GCC CCC TGG CCC CCG CCG GAA 96  
 Leu Glu Ala Pro Ala Pro Leu Glu Glu Ala Pro Trp Pro Pro Pro Glu  
 25 20 25 30

GGG GCC TTC GTG GGC TTC GTC CTC TCC CGC CCC GAG CCC ATG TGG GCG 144  
 Gly Ala Phe Val Gly Phe Val Leu Ser Arg Pro Glu Pro Met Trp Ala  
 35 40 45

	GAG CTT AAA GCC CTG GCC GCC TGC AGG GAC GGC CGG GTG CAC CGG GCA Glu Leu Lys Ala Leu Ala Ala Cys Arg Asp Gly Arg Val His Arg Ala	192
	50 55 60	
5	GCA GAC CCC TTG GCG GGG CTA AAG GAC CTC AAG GAG GTC CGG GGC CTC Ala Asp Pro Leu Ala Gly Leu Lys Asp Leu Lys Glu Val Arg Gly Leu	240
	65 70 75 80	
	CTC GCC AAG GAC CTC GCC GTC TTG GCC TCG AGG GAG GGG CTA GAC CTC Leu Ala Lys Asp Leu Ala Val Leu Ala Ser Arg Glu Gly Leu Asp Leu	288
	85 90 95	
10	GTG CCC GGG GAC GAC CCC ATG CTC CTC GCC TAC CTC CTG GAC CCC TCC Val Pro Gly Asp Asp Pro Met Leu Leu Ala Tyr Leu Leu Asp Pro Ser	336
	100 105 110	
15	AAC ACC ACC CCC GAG GGG GTG GCG CGG CGC TAC GGG GGG GAG TGG ACC Asn Thr Thr Pro Glu Gly Val Ala Arg Arg Tyr Gly Gly Glu Trp Thr	384
	115 120 125	
	GAG GAC GCC GCC CAC CGG GCC CTC CTC TCG GAG AGG CTC CAT CGG AAC Glu Asp Ala Ala His Arg Ala Leu Leu Ser Glu Arg Leu His Arg Asn	432
	130 135 140	
20	CTC CTT AAG CGC CTC GAG GGG GAG GAG AAG CTC CTT TGG CTC TAC CAC Leu Leu Lys Arg Leu Glu Gly Glu Glu Lys Leu Leu Trp Leu Tyr His	480
	145 150 155 160	
	GAG GTG GAA AAG CCC CTC TCC CGG GTC CTG GCC CAC ATG GAG GCC ACC Glu Val Glu Lys Pro Leu Ser Arg Val Leu Ala His Met Glu Ala Thr	528
	165 170 175	
25	GGG GTA CGG CTG GAC GTG GCC TAC CTT CAG GCC CTT TCC CTG GAG CTT Gly Val Arg Leu Asp Val Ala Tyr Leu Gln Ala Leu Ser Leu Glu Leu	576
	180 185 190	
30	GCG GAG GAG ATC CGC CGC CTC GAG GAG GAG GTC TTC CGC TTG GCG GGC Ala Glu Glu Ile Arg Arg Leu Glu Glu Glu Val Phe Arg Leu Ala Gly	624
	195 200 205	
	CAC CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA AGG GTG CTC TTT His Pro Phe Asn Leu Asn Ser Arg Asp Gln Leu Glu Arg Val Leu Phe	672
	210 215 220	
35	GAC GAG CTT AGG CTT CCC GCC TTG GGG AAG ACG CAA AAG ACA GGC AAG Asp Glu Leu Arg Leu Pro Ala Leu Gly Lys Thr Gln Lys Thr Gly Lys	720
	225 230 235 240	
	GCG TCC ACC AGC GCC GCG GTG CTG GAG GCC CTA CGG GAG GCC CAC CCC Arg Ser Thr Ser Ala Ala Val Leu Glu Ala Leu Arg Glu Ala His Pro	768
	245 250 255	
40	ATC GTG GAG AAG ATC CTC CAG CAC CGG GAG CTC ACC AAG CTC AAG AAC	816



	Ile Val Glu Lys Ile Leu Gln His Arg Glu Leu Thr Lys Leu Lys Asn	
	260 265 270	
	ACC TAC GTG GAC CCC CTC CCA AGC CTC GTC CAC CCG AGG ACG GGC CGC	864
5	Thr Tyr Val Asp Pro Leu Pro Ser Leu Val His Pro Arg Thr Gly Arg	
	275 280 285	
	CTC CAC ACC CGC TTC AAC CAG ACG GCC ACG GCC ACG GGG AGG CTT AGT	912
	Leu His Thr Arg Phe Asn Gln Thr Ala Thr Ala Thr Gly Arg Leu Ser	
	290 295 300	
10	AGC TCC GAC CCC AAC CTG CAG AAC ATC CCC GTC CGC ACC CCC TTG GGC	960
	Ser Ser Asp Pro Asn Leu Gln Asn Ile Pro Val Arg Thr Pro Leu Gly	
	305 310 315 320	
	CAG AGG ATC CGC CGG GCC TTC GTG GCC GAG GCG GGT TGG GCG TTG GTG	1008
	Gln Arg Ile Arg Arg Ala Phe Val Ala Glu Ala Gly Trp Ala Leu Val	
	325 330 335	
15	GCC CTG GAC TAT AGC CAG ATA GAG CTC CGC GTC CTC GCC CAC CTC TCC	1056
	Ala Leu Asp Tyr Ser Gln Ile Glu Leu Arg Val Leu Ala His Leu Ser	
	340 345 350	
	GGG GAC GAA AAC CTG ATC AGG GTC TTC CAG GAG GGG AAG GAC ATC CAC	1104
20	Gly Asp Glu Asn Leu Ile Arg Val Phe Gln Glu Gly Lys Asp Ile His	
	355 360 365	
	ACC CAG ACC GCA AGC TGG ATG TTC GGC GTC CCC CCG GAG GCC GTG GAC	1152
	Thr Gln Thr Ala Ser Trp Met Phe Gly Val Pro Pro Glu Ala Val Asp	
	370 375 380	
25	CCC CTG ATG CGC CGG GCG GCC AAG ACG GTG AAC TAC GGC GTC CTC TAC	1200
	Pro Leu Met Arg Arg Ala Ala Lys Thr Val Asn Tyr Gly Val Leu Tyr	
	385 390 395 400	
	GGC ATG TCC GCC CAT AGG CTC TCC CAG GAG CTA GCC ATC CCC TAC GAA	1248
	Gly Met Ser Ala His Arg Leu Ser Gln Glu Leu Ala Ile Pro Tyr Glu	
	405 410 415	
30	GAA GCG GTG GCC TTT ATA GAG CGC TAC TTC CAA AGC TTC CCC AAG GTG	1296
	Glu Ala Val Ala Phe Ile Glu Arg Tyr Phe Gln Ser Phe Pro Lys Val	
	420 425 430	
	CGG GCC TGG ATA GAA AAG ACC CTG GAG GAG GGG AGG AAG CGG GGC TAC	1344
35	Arg Ala Trp Ile Glu Lys Thr Leu Glu Glu Gly Arg Lys Arg Gly Tyr	
	435 440 445	
	GTG GAA ACC CTC TTC GGA AGA AGG CGC TAC GTG CCC GAC CTC AAC GCC	1392
	Val Glu Thr Leu Phe Gly Arg Arg Arg Tyr Val Pro Asp Leu Asn Ala	
	450 455 460	
40	CGG GTG AAG AGC GTC AGG GAG GCC GCG GAG CGC ATG GCC TTC AAC ATG	1440
	Arg Val Lys Ser Val Arg Glu Ala Ala Glu Arg Met Ala Phe Asn Met	

40

	465		470		475		480	
	CCC GTC CAG GGC ACC GCC GCC GAC CTC ATG AAG CTC GCC ATG GTG AAG							1488
	Pro Val Gln Gly Thr Ala Ala Asp Leu Met Lys Leu Ala Met Val Lys							
		485		490		495		
5	CTC TTC CCC CGC CTC CGG GAG ATG GGG GCC CGC ATG CTC CTC CAG GTC							1536
	Leu Phe Pro Arg Leu Arg Glu Met Gly Ala Arg Met Leu Leu Gln Val							
		500		505		510		
	CAC GAC GAG CTC CTC CTG GAG GCC CCC CAA GCG CGG GCC GAG GAG GTG							1584
	His Asp Glu Leu Leu Leu Glu Ala Pro Glu Ala Arg Ala Glu Glu Val							
10		515		520		525		
	GCG GCT TTG GCC AAG GAG GCC ATG GAG AAG GCC TAT CCC CTC GCC GTG							1632
	Ala Ala Leu Ala Lys Glu Ala Met Glu Lys Ala Tyr Pro Leu Ala Val							
		530		535		540		
	CCC CTG GAG GTG GAG GTG GGG ATG GGG GAG GAC TGG CTT TCC GCC AAG							1680
15	Pro Leu Glu Val Glu Val Gly Met Gly Glu Asp Trp Leu Ser Ala Lys							
		545		550		555		560
	GGT TAG							1686
	Gly *							

Claims

1. An enzymatically active DNA polymerase having between 540 and 582 amino acids having a tyrosine at a position equivalent to position 667 of Taq DNA  
5 polymerase, wherein said polymerase lacks 5' to 3' exonuclease activity, and wherein said polymerase has at least 95% homology in its amino acid sequence to the DNA polymerase of Thermus aquaticus, Thermus flavus or  
10 Thermus thermophilus, and wherein said polymerase forms a single polypeptide band or an SDS polyacrylamide gel.
2. The polymerase of claim 1 wherein the amino acid sequence of said polymerase includes less than 3 conservative amino acid changes compared to one said DNA polymerase of said named Thermus species.
- 15 3. The polymerase of claim 1 wherein the amino acid sequence of said polymerase includes less than 3 additional amino acids compared to one said DNA polymerase of said named Thermus species at its N-terminus.
- 20 4. The polymerase of claim 1 selected from the group consisting of FY2, FY3 and FY4.
5. Purified nucleic acid encoding the DNA polymerase of any of claims 1-4.

6. Method for sequencing DNA comprising the step of generating chain terminated fragments from the DNA template to be sequenced with a DNA polymerase of any of claims 1-4 in the presence of at least one chain  
5 terminating agent and one or more nucleotide triphosphates, and determining the sequence of said DNA from the sizes of said fragments.

7. Kit for sequencing DNA comprising a DNA polymerase of any of claims 1-4 and a pyrophosphatase.

- 10 8. The kit of claim 7 wherein said pyrophosphatase is thermostable.

is thermostable?

is thermostable.

9. Apparatus for DNA sequencing having a reactor comprising a DNA polymerase of any of claims 1-4 and a band separator.

Fig. 1A

1081/361 ttc cag gag ggg cgg gac atc cac acg gag acc gcc agc tgg atg ttc tgc gtc ccc cgg gag gcc gtg gac ccc ctg atg atg cgc cgg gcg  
 F Q E G R D I H T E T A S W M F G V P R E A V D P L M R R A  
 1171/391 gcc aag acc atc aac tac ggg gtc ctc tac gcc atg tcg gcc cac cgc ctc tcc cag gag cia gcc atc cct tac gag gag gcc cag gcc  
 A K T I N Y G V L Y G M S A H R L S Q E L A I P Y E A Q A  
 1261/421 ttc att gag cgc tac ttt cag agc ttc ccc aag gtg cgg gcc tgg att gag aag acc ctg gag gag gcc agg agg cgg ggg tac gtg gag  
 F I E R Y F Q S F P K V R A W I E K T L E F G R R G Y V E  
 1351/451 acc ctg ttc gcc cgc cgc tac gtg cca gac cta gag gcc cgg gtg aag agc gtg cgg gag gcg gcc gag cgc atg gcc ttc aac atg  
 T L F G R R R Y V P D L E A R V K S V R E A A E R M A F N M  
 1441/481 ccc gtc cag ggc acc gcc gcc ctc atg aag ctg gct atg gtg aag ctc ttc ccc ccc agg ctg gag gaa atg ggg gcc agg atg ctc ctt  
 P V Q G T A A D L M K L A M V K L F P R L E E M G A R M L L  
 1531/511 cag gtc cac gac gag ctg gtc ctc gag gcc cca aaa gag aag ggc gag gcc ctg gcc cgg ctg gcc aag gag gtc atg gag ggg gtg tat  
 Q V H D E L V L E A P K E R A E A V A R L A K E V M E G V Y  
 1621/541 ccc ctg gcc gtg ccc ctg gag gtg gag ggg ata ggg gag gac tgg ctc tcc gcc aag gag tga  
 P L A V P L E V E V G I G E D W L S A K E \*

Fig. 1B

1/1  
 atg gct ctg gaa cgt ctg gag ttt ggc agc ctc cac gag ttc ggc ctt ctg gaa agc  
 M A L E R L E F G S L L H E F G L L E S P K A L E E A P W P  
 91/31  
 ccg ccg gaa ggg gcc ttc gtg ggc ttt gtg ctt tcc cgc aag gag ccc atg tgg gcc gat  
 P P E G A F V L S R K E P M W A D L L A L A A A A R G G  
 181/61  
 ccg gtc cac cgg gcc ccc gag cct tat aaa gcc ctc agc gag cta ggc cgg ggg  
 R V H R A P E P Y K A L L R D L K E A R G L L A K D L S V L A  
 271/91  
 ctg agg gaa ggc ctt ggc ctc ccg gcc ggc gac gac ccc atg ctc ctc ctg gac cct  
 L R E G L G L P P G D D P M L L A Y L L D P S N T T P E G V  
 361/121  
 gcc c99 gcc tac ggc ggg gag tgg acg gag ggc ggc ggc ggc ggc ggc ggc ggc  
 A R Y G G E W T E A G ggg gag cgg gcc ctt tcc gag agc ctc ttc gcc aac ctg tgg ggg agg ctt  
 451/151  
 gag ggg gag agg ctc ctt tgg ctt tac c99 gag gtg gag agg ccc ctt tcc gct gtc  
 E G R L L W L Y R E V E R P L S A V L A H M E A T G G V R  
 541/181  
 ctg gac gtg gcc tat ctc agg gcc ttg tcc ctg gag gtg ggc gag gag atc gcc cgc  
 L D V A Y L R A L S L E V A E I A R L E A E V F R L A G H  
 631/211  
 ccc ttc aac ctc aac tcc c99 gac cag ctg gaa agg gtc ctc ttt gac gag cta ggg  
 P F N L N S R D Q L L E R V L F D E L G L P A I G K T E K T G  
 721/241  
 aag cgc tcc acc agc gcc gtc ctg gag gcc ctc cgc gag gcc cac ccc atc gtg  
 K R S T S A A V L E A L R E A H P I V E K I I A A T C L Q Y R E L T K  
 811/271  
 ctg aag agc acc att gag ccc ttg ccg gac ctc atc cac ccc agg acg ggc cgc  
 L K S T Y I D P L P D L I H P R T G G R L H T R F N Q T A T A  
 901/301  
 ccg gcc agg cta agt agc tcc gat ccc aac ctc gag aac atc ccc gtc cgc ctt  
 T G R L S S S D P N L Q N I P V R T P L G Q R I R R A F I A  
 991/331  
 gag gag ggg tgg cta ttg gtg gcc ctg gat agc cag ata gag ctc agg gtg ctg  
 E E G W L L V A L D Y S Q I E L R V L A H L S G D E N L I R  
 161/21  
 ggc ccc aag gcc ctg gag gag gcc ccc tgg ccc  
 P K A L E E A P W P  
 151/51  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 241/81  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 331/111  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 421/141  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 511/171  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 601/201  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 691/231  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 781/261  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 871/291  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 961/321  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G  
 1051/351  
 ggc ccc aag gcc ctg gag gag gcc agg ggg ggc  
 L L A L A A A R G G

Fig. 2A

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4 / 1 2

1081/361 gtc ttc cag gag cgg gac atc cac acg gag acc gcc agc tgg atg ttc ggc gtc ccc cgg gag gcc gtg gac ccc ctg atg cgc cgg  
 1171/391 ggc gcc aag acc atc aac tac ggg gtc ctc tac ggc atg tgc gcc cac cgc ctc tcc cag gag cta gcc atc cct tac gag gag gcc cag  
 1261/421 gtc ttc att gag cgc tac ttt cag agc ttc ccc aag gtg cgg gcc tgg att gag aag acc ctg gag gag ggc agg agg cgg ggg tac gtg  
 1351/451 gag acc ctc ttc ggc cgc cgc cgc tac gtg cca gac cta gag gcc cgg gtg aag agc gtg cgg gag gcg gcc gag cgc atg gcc ttc aac  
 1441/481 atg ccc gtc cag ggc acc gcc gac ctc atg aag ctg gct atg gtg aag ctc ttc ccc agg ctg gag gaa atg ggg gcc agg atg ctc  
 1531/511 ctt cag gtc cac gag cgc ctc gtc ctc gag gcc cca aaa gag agg gcg gag gcc gtg gcc cgc cgc atg ggc atg gag ggg gtg  
 1621/541 tat ccc ctg gcc ctg ccc ctg gag gtg gag ggc ggg ata ggg gag gac tgg ctc tcc gcc aag gag tga  
 1111/371 gtc ttc cag gag cgg gac atc cac acg gag acc gcc agc tgg atg ttc ggc gtc ccc cgg gag gcc gtg gac ccc ctg atg cgc cgg  
 1201/401 ggc gcc aag acc atc aac tac ggg gtc ctc tac ggc atg tgc gcc cac cgc ctc tcc cag gag cta gcc atc cct tac gag gag gcc cag  
 1291/431 gtc ttc att gag cgc tac ttt cag agc ttc ccc aag gtg cgg gcc tgg att gag aag acc ctg gag gag ggc agg agg cgg ggg tac gtg  
 1381/461 gag acc ctc ttc ggc cgc cgc cgc tac gtg cca gac cta gag gcc cgg gtg aag agc gtg cgg gag gcg gcc gag cgc atg gcc ttc aac  
 1471/491 atg ccc gtc cag ggc acc gcc gac ctc atg aag ctg gct atg gtg aag ctc ttc ccc agg ctg gag gaa atg ggg gcc agg atg ctc  
 1561/521 ctt cag gtc cac gag cgc ctc gtc ctc gag gcc cca aaa gag agg gcg gag gcc gtg gcc cgc cgc atg ggc atg gag ggg gtg  
 1651/551 tat ccc ctg gcc ctg ccc ctg gag gtg gag ggc ggg ata ggg gag gac tgg ctc tcc gcc aag gag tga

Fig. 2B







7 / 1 2

2161/721 acc ctc ttc ggc cgc cgc tat gtg ccc gac ctc aac gcc cgc cgc aag agc gtg cgc gag gcg gcg gag cgc atg gcc ttc aac atg  
 T L F G R R R Y V P D L N A R V K S V R E A A 2221/741  
 2251/751 ccg gtc cag ggc acc gcc gac ctc atg aag ctg gcc atg gtg cgc ctt ttc ccc cgc ctt cag gaa ctg ggc gcg agg atg ctt ttg  
 P V Q G T A A D L M K L A M V R L F P R L Q E L G A R M L L 2311/771  
 2341/781 gag gtg cac gac gag ctg gtc ctc gag gcc ccc aag gac cgc gcg gag agg gta gcc gct ttg gcc aag gag gtc atg gag ggc gtc tgg  
 Q V H D E L V L E A P K D R A E R V A A L A K E V M E G V W 2401/801  
 2431/811 ccc ctg cag gtg ccc ctg gag gtg gag gac tgg ctc tcc gcc aag gag tag  
 P L Q V P L E V E V G L G E D W L S A K E 2491/831

Fig. 3C

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[illegible]

Fig. 4A

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1081/361 TTT GCC TCG AGG GAG GGG CTA GAC CTC GTG CCC GGG GAC GAC CCC ATG CTC CTC GCC TAC CTC CTC GAC CCC TCC AAC ACC ACC CCC GAG  
 L A S R E G L D L V P G D 1111/371  
 1171/391 GGG GTG GCG CGG CGC TAC GGG GGG GAG TGG AGG GAG GAC GGC GCC CAC CGG GCG CTC CTC TCG GAG AGG CTC CAT CGG AAC CTC CTT AAG  
 G V A R R Y G G E W T E D A A H R A L L S E R L H R N L L K  
 1261/421 CGC CTC GAG GGG GAG AAG CTC CTT TGG CTC TAC CAC GAG GTG GAA AAG CCC CTC TCC CGG GTC CTG GCC CAC ATG GAG GCC ACC GGG  
 R L E G E K L L W L Y H L S L E L A E I R R L E E V F R L A  
 1351/451 GTA CGG CTG GAC GTG GCC TAC CTT CAG GCC CTT TCC CTG GAG CTT GCG GAG ATC CGC CGC CTC GAG GAG GTC TTC CGC TTG GCG  
 V R L D V A Y L Q A L S L E L A E I R R L E E V F R L A  
 1441/481 CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA AGG GTG CTC TTT GAC GAG CTT AGG CTT CCC GCC TTG GGG AAG ACG CAA AAG  
 G H P F N L N S R D Q L E R V L F D E L R L P A L G K T Q K  
 1531/511 ACA GGC AAG CGC TCC ACC AGC GCC GCG GTG CTG GAG GGC CTA CGG GAG GCC CAC CGG AGG AGC GGC CTC CAC CGC CTC CAC ACC CGC TTC AAC CAG ACG GCC  
 T G K L K N T Y V D L P S L V H P R T G R L H T R F N Q T A  
 1621/541 ACC AAG CTC AAG AAC ACC TAC GTG GAC CCC CTC CCA AGC CTC GTG CAC CGG AGG ACG GGC CGC CTC CAC ACC CGC TTC AAC CAG ACG GCC  
 T K L K N T Y V D L P S L V H P R T G R L H T R F N Q T A  
 1711/571 ACG GCC ACG GGG AGG CTT AGT AGC TCC GAC CCC AAC CTG CAG AAC ATC CCC GTC CTC CGC ACC CCC TTG GGC CAG AGG ATC CGC CGG GCC TTC  
 T A T G R L S S D P N L Q N I P V R T P L G Q R I R R A F  
 1801/601 GTG GCC GAG GCG GGT TGG GCG TTG GTG GCC CTG GAC TAT AGC CAG ATA GAG CTC CGC GTC CTC GCC CAC CTC TCC GGG GAC GAA AAC CTG  
 V A E A G W A L V A L D Y S Q I E L R V L A H L S G D E N L  
 1891/631 ATC AGG GTC TTC CAG GAG GGG AAG GAC ATC CAC ACC CAG ACC GCA AGC TGG ATG TTC GGC CTC CCC CGG GAG GCC GTG GAC CCC CTG ATG  
 I R V F Q E G K D I H T Q T A S W M F G V P P E A V D P L M  
 1981/661 CGC CGG GCG GCC AAG ACG GTG AAC TTC GGC CTC TAC GGC ATG TCC GCC CAT AGG CTC TCC CAG GAG CTT GCC ATC CCC TAC GAG GAG  
 R R A A K T V N F G V L Y G M S A H R L S O E L A I P Y E E  
 2071/691 GCG GTG GCC TTT ATA GAG CGC TAC TTC CAA AGC TTC CCC AAC GTG CGG GCC TGG ATA GAA AAG ACC CTG GAG GAG GGG AGG AAG CGG GCG  
 A V A F I E R Y F Q S F P K V R A W I E K T L E E G R K R G

Fig. 4B

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2161/721 TAC GTG GAA ACC CTC TTC GGA AGA AGG CGC TAC GTG CCC GAC CTC AAC GCC CGG GTG AAG AGC GTC AGG GAG GCC GCG GAG CGC ATG GCC  
 Y V E T L F G R R Y V P D L N A R V K S V R E A A E R M A  
 2251/751 TTC AAC ATG CCC GTG CAG GGC ACC GCC GGC GAC CTC ATG AAG CTC GCC ATG GTG AAG CTC TTC CCC CGC CTC CGG GAG ATG GGG GCC CGC  
 F N M P V Q G T A A D L M K L A M V K L F P R L R E M G A R  
 2341/781 ATG CTC CTC CAG GTG CAC GAC GAG CTC CTC CTG GAG GCC CCC CAA GCG CGG GCC GAG GAG GTG GCG GCT TTG GCC AAG GAG GCC ATG GAG  
 M L L Q V H D E L L L E A P Q A R A E E V A A L A K E A M E  
 2431/811 AAG GCC TAT CCC CTC GCC GTG CCC CTC GAG GTG GAG GTG GGG ATG GGG GAG GAC TGG CTT TCC GCC AAG GGT TAG  
 K A Y P L A V P L E V E V G M G E D W L S A K G \*

Fig. 4C



1081/361 TTC CAG GAG GGG AAG GAC ATC CAC ACC CAG ACC GCA AGC TGG ATG TTC GGC GTC CCC CCG GAG GCC GTG GAC CCC CTG ATG CGC CGG GCG  
 F Q E G K D I H T Q T A S W M F G V P P E A V D P L M R R A  
 1111/371  
 1171/391 GCC AAG ACG GTG AAC TAC GGC GTC CTC TAC GGC ATG TCC GGC CAT AGG CTC TCC CAG GAG CTA GCC ATC CCC TAC GAA GAA GCG GTG GCC  
 A K T V N Y G V L Y G M S A H R L S Q E L A I P Y E E A V A  
 1201/401  
 1261/421 TTT ATA GAG CGC TAC TTC CAA AGC TTC CCC AAG GTG CGG GCC TGG ATA GAA AAG ACC CTG GAG GAG GGG AGG AAG CGG GGC TAC GTG GAA  
 F I E R Y F Q S F P K V R A W I E K T L E G R K R G Y V E  
 1291/431  
 1351/451 ACC CTC TTC GGA AGA AGG CGC TAC GTG CCC GAC CTC AAC GCC CGG GTG AAG AGC GTC AGG GAG GCC GCG GAG CGC ATG GCC TTC AAC ATG  
 T L F G R R Y V P D L N A R V K S V R E A A E R M A F N M  
 1381/461  
 1441/481 CCC GTC CAG GGC ACC GCC GAC CTC ATG AAG CTC GCC ATG GTG AAG CTC TTC CCC CCG CTC CGG GAG ATG GGG GCC CGC ATG CTC CTC  
 P V Q G T A A D L M K L A M V K L F P R L R E M G A R M L L  
 1471/491  
 1531/511 CAG GTC CAC GAC GAG CTC CTC CTG GAG GCC CCC CAA GCG CGG GCC GAG GAG GTG GCG GCT TTG GCC AAG GAG GCC ATG GAG AAG GCC TAT  
 Q V H D E L L L E A P Q A R A E E V A A L A K E A M E K A Y  
 1561/521  
 1621/541 CCC CTC GCC GTG CCC CTG GAG GTG GAG GTG GGG ATG GGG GAG GAC TGG CTT TCC GCC AAG GGT TAG  
 P L A V P L E V E V G M G E D W L S A K K G G T \*  
 1651/551  
 1681/561

Fig. 5B



## INTERNATIONAL SEARCH REPORT

International Application No.  
PCT/US 96/06906

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/54 C12N9/12 C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO,A,92 06188 (BARNES WAYNE M) 16 April 1992 cited in the application see the whole document ---	1-9
A	WO,A,91 09944 (CETUS CORP) 11 July 1991 see the whole document ---	1-9
A	WO,A,94 05797 (KISELEV VSEVOLOD ;SEVERIN EVGENII (RU); KORPELA TIMO (FI)) 17 March 1994 see the whole document --- -/-	1-9

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

1 August 1996

Date of mailing of the international search report

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## INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 96/06906

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